

Ultraviolet A and melanoma: A review

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The incidence and mortality rates of melanoma have risen for many decades in the United States. Increased exposure to ultraviolet (UV) radiation is generally considered to be responsible. Sunburns, a measure of excess sun exposure, have been identified as a risk factor for the development of melanoma. Because sunburns are primarily due to UVB (280-320 nm) radiation, UVB has been implicated as a potential contributing factor to the pathogenesis of melanoma. The adverse role of UVA (320-400 nm) in this regard is less well studied, and currently there is a great deal of controversy regarding the relationship between UVA exposure and the development of melanoma. This article reviews evidence in the English-language literature that surrounds the controversy concerning a possible role for UVA in the origin of melanoma. Our search found that UVA causes DNA damage via photosensitized reactions that result in the production of oxygen radical species. UVA can induce mutations in various cultured cell lines. Furthermore, in two animal models, the hybrid *Xiphophorus* fish and the opossum (*Mondelphis domestica*), melanomas and melanoma precursors can be induced with UVA. UVA radiation has been reported to produce immunosuppression in laboratory animals and in humans. Some epidemiologic studies have reported an increase in melanomas in users of sunbeds and sunscreens and in patients exposed to psoralen and UVA (PUVA) therapy. There is basic scientific evidence of the harmful effects of UVA on DNA, cells and animals. Collectively, these data suggest a potential role for UVA in the pathogenesis of melanoma. To date evidence from epidemiologic studies and clinical observations are inconclusive but seem to be consistent with this hypothesis. Additional research on the possible role of UVA in the pathogenesis of melanoma is required. (J Am Acad Dermatol 2001;44:837-46.)

The incidence and mortality rates of melanoma have been rising during the past several decades in the United States.^{1,2} Among the reasons for these trends, increased exposure to ultraviolet (UV) radiation, as a result of lifestyle changes, has been generally recognized as an important factor. Sunburns, a measure of excessive sun

Abbreviations used:

IPD:	immediate pigment darkening
IPD-PF:	immediate pigment darkening protection factor
MED:	minimal erythema dose
PPD:	persistent pigment darkening
PUVA:	psoralen (P) and ultraviolet A (UVA)
SPF:	sun protection factor
UV:	ultraviolet
UVA:	ultraviolet A
UVB:	ultraviolet B

exposure, have been determined to be a risk factor for the development of melanoma. Because sunburns are primarily caused by UVB (280-320 nm) radiation and UVB is strongly absorbed by DNA, which results in chromosomal damage, it has been inferred that this portion of the solar UV spectrum could be a major causative factor for melanoma. However, the adverse role of UVA (320-400 nm) as a possible etiologic factor for melanoma has received less attention, even though excessive sun exposure

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is also associated with excessive exposure to UVA (320-400 nm).

The UVA bandwidth (80 nm) is twice that of UVB (40 nm), and 90% to 95% of solar UV radiation energy (measured in watts per square meter) that reaches the surface of the earth is UVA³; only 5% to 10% of it is UVB. In addition to solar radiation, high-dose exposure to UVA can also come from devices such as sunbeds/sunlamps and PUVA therapy units. UVA has longer wavelengths than UVB and thus penetrates deeper into skin. It is estimated that about 19% to 50% of the solar UVA can reach the depth of melanocytes,⁴⁻⁶ whereas only about 9% to 14% of solar UVB reaches the melanocytes.⁴⁻⁶ This long-wavelength property of UVA also allows it to pass through most automobile, office, and household windows, whereas UVB is blocked by window glass. Recently, during a study on UV protection offered by cotton fabrics used in summer clothing,⁷ a substantially greater percent transmission of UVA, as compared with UVB, occurred through these fabrics. This means that people wearing summer clothing may receive substantially more UVA than UVB even in clothed areas.

The two UV spectra also have different biologic effects. UVB is about 1000 times more effective than UVA in inducing sunburn.⁸ UVA, on the other hand, is much more potent in inducing immediate pigment darkening (IPD)^{9,10} and persistent pigment darkening (PPD).¹¹ Both types of pigment darkening are characterized by darkening of the skin immediately after UVA exposure. The observed pigment in IPD fades within minutes after exposure, whereas PPD is stable for at least 2 to 3 hours. The induction of IPD and PPD is readily observed in dark-skinned persons (Fitzpatrick type III and above) and more difficult to see in those with light skin (Fitzpatrick types I and II). Because UVA does not readily produce erythema or pigment darkening in light-skinned persons, the immediate effects after UVA exposure in light-skinned persons are virtually "silent." However, biologic damage occurs. UVA causes formation of singlet oxygen radicals that subsequently lead to DNA-strand breakage, nuclear base damage, and mutations.

The possibility that UVA can cause melanoma in humans has been hypothesized by Garland, Garland, and Gorham.¹² They suggested that the rising incidence of melanoma might in part be related to the widespread use of sunscreens providing only UVB protection, thereby allowing more prolonged exposure to UVA radiation. In addition, Setlow et al¹³ demonstrated that UVA is capable of producing melanoma in *Xiphophorus* hybrid fish.

Currently, the role of UVA in the pathogenesis of melanoma is controversial. In this study, we review

evidence from the basic sciences, epidemiology, and clinical observations concerning this controversy. The role of UVB in causing melanoma is not discussed in this review.

BASIC SCIENCE EVIDENCE

Cancer may be viewed as a disease that progresses through multiple stages that involve initiation, promotion, progression, and metastasis. Each stage is accompanied by further changes in DNA. UVB is known to cause mutations in oncogenes and tumor suppressor genes that eventually initiate the cascade of events resulting in skin cancers.¹⁴⁻¹⁶ To substantiate that UVA has the potential to induce melanoma, it is necessary to demonstrate that UVA is capable of producing biologic damage. It has been shown that UVA is capable of producing DNA damage via photosensitized reactions that result in oxygen radical species (eg, hydrogen peroxide, singlet oxygen, superoxide anion).¹⁷ These reactive species can cause single-strand breaks,¹⁸ mutations, sister chromatid exchanges, and chromosomal aberrations that can result in cytotoxicity and carcinogenesis.¹⁹ Furthermore, the singlet oxygen and superoxide anions generated by UVA1 (340-400 nm) can damage mitochondria and induce apoptosis in cell culture.²⁰

UVA damage to DNA in cell-culture models

UVA radiation has been shown to cause mutations in mammalian cells. Studies in murine cell lines have demonstrated that broadband UVA can induce mutations.^{21,22} In addition, monochromatic UVA at 365 and 334 nm are mutagenic to a human epithelial P3 cell line²³ and to a Chinese hamster ovary cell line,²⁴ respectively. Enninga et al²⁵ have demonstrated that radiation at 365 nm is potentially mutagenic to cultured human fibroblasts. Wenczel et al²⁶ measured DNA single-strand breaks in cultured human melanocytes after UVA exposure. They showed that pheomelanin or melanin intermediates (or both) were the most likely chromophores that react with the UVA radiation, leading to the DNA single-strand breaks.

Using monochromatic radiation, Kvam and Tyrrell²⁷ demonstrated that the wavelengths causing almost all of the oxidative DNA base damage (eg, 7,8-dihydro-8-oxoguanine) in a human skin fibroblast cell line are in the UVA and visible light range. In addition, they estimated that the total amount of such guanine base damage induced by sunlight in fibroblasts of the skin equals or exceeds the amount of the major type of direct DNA damage, cyclobutane pyrimidine dimers induced mainly by UVB. Drobetsky, Turcotte, and Chateaufort²⁸ have characterized a specific mutation at the ade-

nine phosphoribosyltransferase locus in Chinese hamster ovary cells irradiated with UVA. They demonstrated a high frequency ($\leq 50\%$) of T \rightarrow G transversion, a rare class of mutations ("fingerprint" mutation), in UVA-irradiated cells, compared with 9% in UVB-irradiated cells. A moderately high frequency (25%) of T \rightarrow G transversion was seen with cells irradiated with simulated sunlight, leading to their conclusion that most of the T \rightarrow G transversion mutations in cells irradiated with simulated sunlight can be attributed to the UVA portion of sunlight, with little, if any, contribution from UVB.

In addition to the aforementioned studies with different mammalian cell lines, the potential carcinogenic effect of UVA has also been demonstrated in cultured human melanocytes. Marrot et al²⁹ induced DNA breaks in the nucleus of Caucasian human melanocytes with broad-spectrum UVA (320-400 nm) irradiation. DNA breakage in cells was assessed via the comet assay, a technique used extensively for analyzing genotoxic effects by environmental chemicals³⁰ or by UV components of the solar spectrum.^{31,32} The investigators suggested that the endogenous pigments and/or melanin-related molecules seem to enhance DNA breakage after UVA irradiation. This is evidenced by higher DNA breakage in melanocytes than in fibroblasts, in cells with higher melanin content, and in cells stimulated for melanogenesis by culturing in tyrosine-rich medium. In addition, Marrot et al²⁹ demonstrated that there is an increased level of p53 expression in the irradiated melanocytes, indicating DNA damage.

That different UVA wavelengths induce mutations in different cell lines can be explained by the different types and amounts of photosensitizers or chromophores in these cells. Unlike UVB, UVA must first react with endogenous photosensitizers (eg, flavins, porphyrins, melanins) that in turn generate reactive oxygen species that cause single-strand breaks or photoadducts. The different sensitivities and mechanisms for DNA damage induced by UVA and by UVB are important to elucidate to improve the formulation of sunscreens with broad-spectrum coverage. Because the most effective wavelength in causing photoproduct formation in human skin is around 300 nm in the UVB spectrum,³³ efforts could be focused on developing UVB-protective sunscreens with maximal protection at or around this wavelength. However, the various photosensitizers in skin have different absorption spectra. These photosensitizers absorb different spectral bands of UVA and then generate oxygen radicals. Thus for UVA-protective sunscreens to be maximally effective, they should have broad coverage across the entire UVA spectrum.

Animal models

Two animal models have demonstrated the possible role of UVA in inducing melanoma and melanoma precursor lesions. Setlow et al¹³ showed that the pigmented backcross hybrids of the genus *Xiphophorus* (platyfish and swordtail) are very sensitive to melanoma induction after a single exposure to any of several UV monochromatic radiations (302, 313, 365, 405, and 436 nm). The wavelength that yielded the highest induction rate per incident photon was 302 nm. However, after factoring in the intensity of various wavelengths in sunlight, it was found that the 365-nm wavelength in sunlight would be the most effective wavelength for melanoma induction in these fish. These authors also suggested that if the action spectrum for human melanoma was similar to that of the fish, then approximately 90% of the melanoma-inducing effects of sunlight might result from UVA and visible radiation.

Ley³⁴ demonstrated that melanoma precursors could be induced in opossums (*Monodelphis domestica*) after prolonged exposure to broad-spectrum UVA. The animals were exposed to 2.5×10^4 J/m² of UVA 3 times per week. After a period of 81 weeks, 22% of the animals developed melanocytic hyperplasia, a precursor of melanoma.

UVA-induced DNA damage in human skin

In an effort to directly assess the potential for UVA to induce DNA damage in human skin, Burren et al³⁵ irradiated the buttocks of healthy volunteers with either UVA1 (340-400 nm) alone, UVA1 plus UVA2 (320-340 nm), or solar-simulating radiation, which contains UVB in addition to UVA. They assayed for both p53 protein expression and the presence of pyrimidine dimers. Pyrimidine dimers are formed in DNA as the result of UV-induced damage; increased p53 levels are part of an effort by cells to repair such damage. Irradiation with UVA1 plus UVA2 induced p53 in epidermal keratinocytes. The levels were about half that observed with equivalent doses of solar-simulated UV. Low doses of UVA alone induced p53 expression in basal keratinocytes, whereas higher doses affected all layers of the epidermis. An erythral dose of solar-simulated UV also induced p53 in all epidermal layers. All 3 types of radiation resulted in similar amounts of pyrimidine dimer formation. These studies provide in vivo evidence that DNA damage in human skin may be caused not only by UVB, but also by UVA.

Role of UVA in inducing immunosuppression

UV radiation, especially UVB, has been implicated in causing immunosuppression.^{36,37} Mainly from work with UVB radiation, it is known that the num-

ber of antigen-presenting Langerhans cells and their functions are adversely affected by UV. Furthermore, UV radiation induces keratinocytes to release various cytokines that influence immune balance in skin. Three cytokines, interleukin 10 (IL-10), IL-12, and interferon gamma (IFN- γ), have been widely studied. IL-10 appears to have an immunosuppressive function, inhibiting antigen-presentation ability of the Langerhans cells³⁸ and suppressing the contact hypersensitivity response.³⁹ On the other hand, IL-12 and IFN- γ have immunopotentiating effects that promote and enhance T helper cell 1 activity. In addition, IFN- γ appears to block the production of IL-10 by macrophages,⁴⁰ possibly mitigating potential IL-10-induced immunosuppression.

There is still a great deal of controversy concerning the effect of UVA in causing immunosuppression. An in vitro study by Iwai et al⁴¹ demonstrated that UVA irradiation decreased the ability of murine epidermal keratinocytes to present antigens to T cells. This decrease was dose-dependent. In addition, they showed that UVA radiation also decreased the expression of costimulatory molecules (ie, intercellular adhesion molecule 1, B7-1, and B7-2) on Langerhans cells. Costimulation is a necessary component for immune action. The antioxidant glutathione was able to mitigate the UVA-induced suppression of antigen-presenting function in a dose-dependent fashion. They concluded that UVA radiation induces immune suppression, at least partially, via an oxidative pathway.

In an animal study using a contact hypersensitivity model, Bestak and Halliday⁴² irradiated C3H/HeJ mice with low-dose UVA (average cumulative dose of 45.9 J/cm²) over a 4-week period. They demonstrated that UVA caused a significant reduction in the number of epidermal Langerhans cells and local but not systemic immunosuppression in these animals. In contrast, Reeve et al⁴³ found that in hairless mice a single dose of UVA exposure (38.7 J/cm²) not only was immunologically innocuous but also reversed the immunosuppressive effect of UVB radiation and *cis*-urocanic acid in a contact-hypersensitivity model.

Evidence for immunosuppression in humans also comes from contact hypersensitivity studies. Using subjects allergic to nickel, Damian, Barnetson, and Halliday⁴⁴ demonstrated that low-dose UVA exposure induced immunosuppression. The UVA-induced immunosuppressive effect was maximal in subjects receiving a 3-day course of UVA exposure, but subjects receiving 4- or 5-day courses of UVA exposure did not show significant immunosuppression. They suggested that acute UVA exposure may initially be immunosuppressive but subsequently be immunoprotective. Potential mechanisms that restore the

immune functions were suggested; these include the formation of antioxidant ferritin in the dermis and the formation of photoadducts with inhibitory effects on *cis*-urocanic acid. LeVee et al⁴⁵ demonstrated that a single exposure to UVA2 (320-340 nm) at 4 minimal erythema doses (MEDs) reduces the number of Langerhans cells. In addition, they have shown that UVA2 leads to decreased local sensitization to dinitrochlorobenzene in human subjects. Furthermore, Fourtanier et al⁴⁶ demonstrated that sunscreens with enhanced UVA protection were more effective in reducing immunosuppression by solar-simulated radiation. However, Sjoval and Christensen⁴⁷ found that UVB, but not UVA, suppressed contact hypersensitivity to nickel antigen in nickel-sensitive persons. The subjects in this study received total body irradiation with a cumulative UVA dose of 205 J/cm² over a 3-week period, a UVA dose that is higher than the subjects received in the studies by Damian, Barnetson, and Halliday⁴⁴ and LeVee.⁴⁵

These conflicting results on the effects of UVA on immunosuppression may be related to the different protocols in these studies, namely: (1) the presence of different amounts of UVB contamination, (2) low versus high UVA dose, and (3) single exposure versus repeated exposures. Further studies are needed to resolve these discrepancies. However, until then, the possibility of UVA-induced immunosuppression and the consequent immune changes that allow the development of melanoma should be considered, especially in view of the fact that melanoma is an immunogenic tumor that appears to respond to immune-based therapy⁴⁸⁻⁵⁰ and the fact that immunosuppressed patients have a higher risk of developing melanoma. Children with immunodeficiency have a 3- to 6-fold increased risk of developing melanoma, and those with Hodgkin's disease have an 8-fold increased risk.⁵¹ Adults with Hodgkin's disease,⁵² and those having kidney or cardiothoracic transplantation,^{53,54} have an increased risk for the development of melanoma.

EPIDEMIOLOGIC EVIDENCE

Epidemiologic observations often reveal associations that provide valuable clues and new channels for further investigations, but these observations are not a substitute for understanding the pathogenesis of diseases. The validity of epidemiologic studies can be undermined by recall bias and confounding factors.

Sunbeds and sunlamps

Every year, approximately 25 million Americans use sunbeds/sunlamps,^{55,56} and this use is particularly prominent in teenagers and young adults. The popularity of tanning salons can be traced largely to

the pervasive belief that tanned skin is more attractive and fashionable than pale skin. In addition, the tanning industry has recommended developing a "safe and protective tan" before vacationing in sunny climates.

Unfortunately, cosmetic indoor tanning has various adverse health effects.⁵⁶ One such effect is that persons using tanning salons receive a large quantity of unnecessary UVA exposure. Currently, most of the tanning lamps on the market emit nearly 100% UVA radiation.^{3,57} In addition, Miller et al³ calculated that a typical tanner (20 sessions at 2 MEDs per session) could receive an annual UVA dose from the sunlamps that is 0.3 to 1.2 times that of an average annual cumulative UVA dose from the sun, which is estimated to be 7700 kJ/m². For more avid tanners (100 sessions at 4 MEDs per session), the UVA dose from sunlamps is 1.2 to 4.7 times that of their annual UVA dose from the sun, which is estimated to be 19,250 kJ/m². The higher estimated annual UVA dose from the sun seen in more avid tanners results from their greater sun-seeking behavior.⁵⁸ The UVA dose is even higher (12 times that from the sun) for the persons in the avid tanner group who use the high-pressure UVA sunlamps.³ If UVA in fact plays a role in the development of melanoma, then the use of sunbeds with primary UVA emission could potentially be a contributor to the development of melanoma.

In 1998, Swerdlow and Weinstock⁵⁵ reviewed 19 case-control studies on the association of the use of tanning lamps and melanoma and reported that 6 of these 19 studies revealed a positive association between the use of tanning lamps and the development of melanoma. In some studies, this association was noted to have a dose-response relationship. However, problems were noted in many of these studies, such as the lack of detailed information on the spectral output of the sunlamps, recall bias, and potential confounding factors. These methodologic issues led the authors to conclude: "Although several investigators have found a positive relation between tanning lamp use and melanoma, in some instances, including dose-response or duration-response effects, the methodologic limitations preclude any firm conclusion regarding a causative relation." Swerdlow and Weinstock urged additional studies to elucidate the relationship between the use of these sunlamps and the development of melanoma.

In 1998, Chen et al,⁵⁹ who studied a population in Connecticut, demonstrated no significant difference overall between persons who ever used a sunlamp and those who never used a sunlamp. However, in persons who first used sunlamps at an age younger than 25 years, they found a borderline significant increase in the risk for the development of

melanoma. The odds ratio for this group was 1.4 (95% confidence interval [CI], 0.9-2.1). One unequivocal finding from this study was that persons who used more than one type of sunlamp were more likely to develop melanoma. The odds ratio was 3.5 (95% CI, 1.3-9.1).

In 1999, Walter, King, and Marrett⁶⁰ reanalyzed their 1985 data to investigate the effects of intermittent and long-term sun exposure in a Canadian population. After adjustment for skin type, age, and sex, the odds ratio for the development of melanoma for those who have used sunbeds and sunlamps, one of the indicators for intermittent exposure, is approximately 1.5 (95% CI, 1.16-2.05). This increased risk is similar to other indicators of intermittent exposure to UV radiation, but higher than for long-term UV exposure. Unfortunately, the precise wavelengths of the sunlamps used by the subjects of this study are not known.

In 2000, through a population-based, matched, case-control study involving 571 patients with melanoma and 914 healthy controls, Westerdahl et al⁶¹ evaluated the association between sunbed/sunlamp use and the development of melanoma in Sweden. After adjusting for hair color, skin type, and number of sunburns, they reported a significantly increased odds ratio for the development of melanoma after regular exposure to sunbeds (odds ratio, 1.8; 95% CI, 1.2-2.7). In addition, they reported a dose relationship between the total number of sunbed uses and melanoma risk. They also reported an odds ratio of 2.3 (95% CI, 1.2-4.2) in regular sunbed users who first used sunbeds before 36 years of age. The majority of the surveyed subjects in the study started to use tanning beds after 1980. Diffey and Farr⁶² have demonstrated that tanning beds from the early 1980s produced mainly UVA plus a small fraction of UVB (<0.1%-2.1%). Hence Westerdahl et al⁶¹ believed that the subjects in their study were mostly exposed to tanning beds that emit mainly UVA. Based on the results of this epidemiologic study and evidences from other studies,^{3,13,63} Westerdahl et al⁶¹ were tempted to suggest that UVA may play a role in the development of melanoma.

The current epidemiologic data raise the issue of a possible relationship between sunlamp use and increased risk for melanoma, but are not conclusive. Additional studies focusing on the relationship between the development of melanoma and the use of sunlamps that emit primarily UVA are needed.

Sunscreen data

Sunburns have been identified as a marker of increased risk of melanoma^{64,65} because of the interaction of high levels of intense intermittent sun

exposure on unadapted, sensitive skin. Sunscreen use is effective in preventing sunburn, actinic keratoses,^{66,67} and squamous cell carcinomas.⁶⁸ For these reasons, it is generally assumed that sunscreens also offer protection against melanoma. Thirteen studies have examined the relationship between sunscreen use and melanoma and found inconsistent results.^{64,69-79} (Berwick et al, unpublished data) Whereas 6 studies demonstrated a statistically significant positive association between sunscreen use and increased risk for the development of melanoma, 3 showed statistically significant inverse association with sunscreen use. Furthermore, 4 studies showed no association between sunscreen use and risk for the development of melanoma. These conflicting findings may partially be explained by the intractable problem of confounding, that is (1) sun-sensitive persons who are inherently predisposed to the development of melanoma are those most likely to use sunscreens, and (2) sunscreen use allows persons to extend their hours of sun exposure.⁸⁰ Measurement error is likely to be large as well because subjects' use of sunscreen is imprecise.

In relation to the scope of this review, the sunscreen data are consistent with a possible role for UVA in the development of melanoma, especially when one considers that earlier sunscreens were developed to protect users from sunburns. Because UVB is 1000 times more effective than UVA in causing sunburn, the original sunscreens were developed to protect against UVB, but not against UVA. Even after the introduction of UVA-absorbing chemicals for sunscreens in 1989, most of these products only provided partial rather than broad-spectrum coverage for UVA. Currently, in the United States, the values of sun protection factor (SPF) on sunscreen products indicate the degree of protection against sunburn. Because the contribution of UVA in causing sunburn is minimal, the protection measurement based on the current SPF standard does not adequately assess UVA protection. Kaidbey and Barnes⁸¹ assessed the ability of several sunscreens with different SPF values to inhibit the IPD reaction, which they called the immediate pigment darkening protection factor (IPD-PF). IPD has been suggested as an end point for assessing protection against UVA because UVA is more potent than UVB in inducing IPD. These authors demonstrated that there is no correlation between SPF values and IPD-PF. In addition, they showed that high SPF sunscreens may have, at best, only a modest IPD-PF. Therefore use of these high SPF sunscreens with relatively low IPD-PF values protect persons against sunburn, allowing the sunscreen users to be exposed to higher doses of UVA by staying in the sun longer. If UVA radiation is even-

tually proven to play a role in the development of melanoma, it follows that the use of sunscreens without adequate UVA protection may have accounted for some of the current increase in melanoma incidence.^{82,83}

Fluorescent light data

Although the initial report of Beral et al⁸⁴ suggested the possible association of exposure to fluorescent lighting (which emits UVA radiation) as a risk factor for melanoma, subsequent reports⁸⁵⁻⁸⁸ have cast doubt on this concept. Further work needs to be done to elucidate the role of exposure to fluorescent lighting as a factor in the causation of melanoma.

CLINICAL EVIDENCE

There are few controlled clinical studies investigating the independent effects that UVA radiation exerts on human skin. However, there are several clinical situations in which UVA is used to treat cutaneous diseases (eg, psoriasis, atopic dermatitis, cutaneous T-cell lymphomas, granuloma annulare, scleroderma)⁸⁹ from which indirect conclusions can be drawn. Unlike therapeutic UVB radiation, therapeutic UVA plus psoralens is associated with the development of large, irregular, unevenly pigmented, dark lentigines called "PUVA lentigines."⁹⁰ These lentigines frequently develop on skin that is not normally exposed to sunlight, such as the genitals. They frequently have a stellate configuration.^{90,91} PUVA lentigines begin appearing 6 to 15 months after the initiation of PUVA therapy.^{91,92} Histologically, there is a proliferation of melanocytes, many of which are large, clustered, binucleate, and atypical.^{91,93-95} The melanocytes of PUVA lentigines often have many long dendrites and show cellular pleomorphism, nuclear hyperchromatism, and angular nuclei.⁹⁶ Ultrastructurally, there is an increase in the number and size of the melanosomes and an increase in DOPA reactivity.^{91,97} There also is an increase in the pigmentation of keratinocytes with supranuclear caps extending about 3 cell layers up from the basal layer.⁹⁶ The atypical melanocytes induced by PUVA can persist for 7 years or more after cessation of PUVA therapy.⁹⁸ Similar atypical lentiginous melanocytic proliferations have also been observed in persons who are not taking psoralens but receive UVA exposure in tanning beds.^{99,100} Finally, PUVA lentigines are clinically and histologically similar to the lentigines found in patients with xeroderma pigmentosum.¹⁰¹

Given the evidence that UVA radiation can stimulate melanocyte proliferation, cause DNA aberrations, and modify gene expression, it seems reasonable to hypothesize that UVA radiation could theoretically increase the risk for the development of

melanoma. Indeed, Stern, Nichols, and Vakeva⁶³ published a study in support of this hypothesis in which they showed that patients treated with PUVA therapy had more than a 5-fold increased relative risk for the development of melanoma. They observed a cohort of 1380 PUVA-treated patients with psoriasis from 1975 to 1997. In 6 of these patients, invasive melanomas developed within the first 15 years after their first PUVA treatment, in 7 patients invasive melanomas developed more than 15 years after their initial PUVA treatment, and in an additional 4 patients in situ melanomas developed. Many of these melanomas appear to have had a latency of as long as 10 to 15 years before becoming clinically apparent. However, Swedish PUVA follow-up studies on bath-PUVA (944 patients)¹⁰² and systemic PUVA (4799 patients)¹⁰³ failed to show an increased risk for the development of melanoma during an average follow-up of between 15 and 16 years. Thus there are clinical data suggesting an association between PUVA therapy and development of melanoma, but the evidence is not conclusive. Furthermore, it has been argued that psoralens are DNA adducts and that the carcinogenic effects of PUVA may result from the mutagenic effects of psoralens after UVA radiation, not a direct effect of UVA.

DISCUSSION

In this review, we have examined evidence from various disciplines that surround the controversy concerning UVA as a causative factor in the development of human melanoma. Although no studies have shown this conclusively, we believe that the evidence currently available to us does not allow this relationship to be ruled out.

The clinical implications of a role for UVA in causing human melanoma are important. People who frequently use sunbeds and sunlamps have exposed themselves to large quantities of unnecessary UVA radiation. Furthermore, a person with a tan can stay in the sun longer before a sunburn develops, thereby receiving even more UVA exposure. As for sunscreen use, it is accepted that sunscreens not only offer protection against sunburn but also against actinic keratoses and squamous cell carcinomas.^{66-68,104,105} However, most early sunscreens provided predominantly UVB but little or no protection against UVA-induced skin damage. Wearing those early sunscreens allows persons to prevent sunburn, to stay out in the sun longer, and inadvertently, to receive larger doses of UVA radiation. Furthermore, the change in UVA intensity with latitude is much smaller than that for UVB, and the change in melanoma incidence with latitude is much smaller than that for squamous cell carcinoma.¹⁰⁶ These data

may support the idea that UVA is important in the induction of melanoma.

Hence, if the hypothesis that UVA plays a role in the pathogenesis of melanoma is valid, the public should be instructed to avoid the use of sunbeds and sunlamps, and when using sunscreens, to use products that provide adequate UVB and broad-spectrum UVA protection. Furthermore, they should also be encouraged to wear clothing for UV protection. Finally, people should be reminded that avoidance of excessive sun exposure is still the best strategy for preventing both UVB- and UVA-induced skin damage. Aside from public education, further research by the sunscreen, fabric, and chemical industries to develop new generations of products that offer better UVB and UVA protection would be helpful.

Effort is also needed to identify an appropriate specific end point for measuring protection against UVA. Currently, there are 3 *in vivo* end points (ie, erythema, IPD, and PPD) for assessing UVA protection being considered. However, the lack of consensus on which end point to use can hamper the development and assessment of UVA protective products. In selecting the appropriate end point for UVA protection, it is important to keep in mind that the ultimate objective of UVA protection is not to inhibit that end point (eg, PPD). Rather, the ideal end point should serve as the closest surrogate for assessing UVA protection against UVA-induced melanocarcinogenesis and immunosuppression. Finally, effort is needed to determine the electromagnetic action spectra for melanoma in humans.

In summary, we have reviewed evidence from various disciplines regarding the controversy concerning the role of UVA in the development of melanoma. It is evident that UVA is capable of inducing DNA damage in cell culture and in humans *in vivo*. UVA also appears to be capable of producing melanoma in backcross hybrids of *Xiphophorus* fish. However, evidence from epidemiologic studies and clinical observations is inconclusive. Further studies are needed to elucidate the scope of the relationship between UVA exposure and development of melanoma. Until a firm conclusion is reached, it is recommended that the public be instructed to minimize their exposure to both UVA and UVB radiation.

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REFERENCES

1. Rigel DS, Friedman RJ, Kopf AW. The incidence of malignant melanoma in the United States: issues as we approach the 21st century. *J Am Acad Dermatol* 1996;34:839-47.

2. Hall HI, Miller DR, Rogers JD, Bewerse B. Update on the incidence and mortality from melanoma in the United States. *J Am Acad Dermatol* 1999;40:35-42.
3. Miller SA, Hamilton SL, Wester UG, Cyr WH. An analysis of UVA emissions from sunlamps and the potential importance for melanoma. *Photochem Photobiol* 1998;68:63-70.
4. Schothorst A, Enniga E, Simons J. Mutagenic effects per erythral dose of artificial and natural sources of ultraviolet light. In: Passchier WF, Bosnjakovic BFM, editors. Human exposure to ultraviolet radiation: risks and regulations; proceedings of a seminar held in Amsterdam, 23-25 March 1987. New York: Excerpta Medica; 1987. p. 103-7.
5. Kaidbey KH, Agin PP, Sayre RM, Kligman AM. Photoprotection by melanin: a comparison of black and Caucasian skin. *J Am Acad Dermatol* 1979;1:249-60.
6. Bruls WA, van Weelden H, van der Leun JC. Transmission of UV-radiation through human epidermal layers as a factor influencing the minimal erythema dose. *Photochem Photobiol* 1984;39:63-7.
7. Wang SQ, Kopf AW, Marx J, Bogdan A, Polsky D, Bart RS. Reduction of ultraviolet transmission through fabrics used in the manufacture of summer wear: clinical implications. *J Am Acad Dermatol* 2001;44:767-74.
8. McKinlay AF, Diffey BL. A reference action spectrum for ultraviolet induced erythema in human skin. *CIE J* 1987;6:17-22.
9. Irwin C, Barnes A, Veres D, Kaidbey K. An ultraviolet radiation action spectrum for immediate pigment darkening. *Photochem Photobiol* 1993;57:504-7.
10. Rosen CF, Jacques SL, Stuart ME, Gange RW. Immediate pigment darkening: visual and reflectance spectrophotometric analysis of action spectrum. *Photochem Photobiol* 1990;51:583-8.
11. Chardon A, Moyal D, Hourseau C. Persistent pigment darkening response as a method for evaluation of ultraviolet A protection assays. In: Lowe NJ, Shaath NA, Pathak MA, editors. Sunscreens: development, evaluation, and regulatory aspects. New York: Marcel Dekker; 1997. p. 559-82.
12. Garland CF, Garland FC, Gorham ED. Rising trends in melanoma: an hypothesis concerning sunscreen effectiveness [see comments]. *Ann Epidemiol* 1993;3:103-10.
13. Setlow RB, Grist E, Thompson K, Woodhead AD. Wavelengths effective in induction of malignant melanoma. *Proc Natl Acad Sci U S A* 1993;90:6666-70.
14. Brash DE, Ziegler A, Jonason AS, Simon JA, Kunala S, Leffell DJ. Sunlight and sunburn in human skin cancer: p53, apoptosis, and tumor promotion. *J Invest Dermatol Symp Proc* 1996;1:136-42.
15. Sarasin A. The molecular pathways of ultraviolet-induced carcinogenesis. *Mutat Res* 1999;428:5-10.
16. van Kranen HJ, de Grujil FR. Mutations in cancer genes of UV-induced skin tumors of hairless mice. *J Epidemiol* 1999;9(Suppl):S58-65.
17. Ananthaswamy HN, Pierceall WE. Molecular mechanisms of ultraviolet radiation carcinogenesis. *Photochem Photobiol* 1990;52:1119-36.
18. Peak JG, Peak MJ. Comparison of initial yields of DNA-to-protein crosslinks and single-strand breaks induced in cultured human cells by far- and near-ultraviolet light, blue light and X-rays. *Mutat Res* 1991;246:187-91.
19. Cerutti PA. Prooxidant states and tumor promotion. *Science* 1985;227:375-81.
20. Godar DE. UVA1 radiation triggers two different final apoptotic pathways. *J Invest Dermatol* 1999;112:3-12.
21. Hitchins VM, Withrow TJ, Olvey KM, Harleston BA, Ellingson OL, Bostrom RG. The cytotoxic and mutagenic effects of UVA radiation on L5178Y mouse lymphoma cells. *Photochem Photobiol* 1986;44:53-7.
22. Lundgren K, Wulf HC. Cytotoxicity and genotoxicity of UVA irradiation in Chinese hamster ovary cells measured by specific locus mutations, sister chromatid exchanges and chromosome aberrations. *Photochem Photobiol* 1988;47:559-63.
23. Jones CA, Huberman E, Cunningham ML, Peak MJ. Mutagenesis and cytotoxicity in human epithelial cells by far- and near-ultraviolet radiations: action spectra. *Radiat Res* 1987;110:244-54.
24. Wells RL, Han A. Action spectra for killing and mutation of Chinese hamster cells exposed to mid- and near-ultraviolet monochromatic light. *Mutat Res* 1984;129:251-8.
25. Enninga IC, Groenendijk RT, Filon AR, van Zeeland AA, Simons JW. The wavelength dependence of u.v.-induced pyrimidine dimer formation, cell killing and mutation induction in human diploid skin fibroblasts. *Carcinogenesis* 1986;7:1829-36.
26. Wenczl E, Van der Schans GP, Roza L, Kolb RM, Timmerman AJ, Smit NP, et al. (Pheo)melanin photosensitizes UVA-induced DNA damage in cultured human melanocytes. *J Invest Dermatol* 1998;111:678-82.
27. Kvam E, Tyrrell RM. Induction of oxidative DNA base damage in human skin cells by UV and near visible radiation. *Carcinogenesis* 1997;18:2379-84.
28. Drobetsky EA, Turcotte J, Chateaufneuf A. A role for ultraviolet A in solar mutagenesis. *Proc Natl Acad Sci U S A* 1995;92:2350-4.
29. Marrot L, Belaidi JP, Meunier JR, Perez P, Agapakis-Causse C. The human melanocyte as a particular target for UVA radiation and an endpoint for photoprotection assessment. *Photochem Photobiol* 1999;69:686-93.
30. Belpaeme K, Cooreman K, Kirsch-Volders M. Development and validation of the in vivo alkaline comet assay for detecting genomic damage in marine flatfish. *Mutat Res* 1998;415:167-84.
31. Arlett CF, Lowe JE, Harcourt SA, Waugh AP, Cole J, Roza L, et al. Hypersensitivity of human lymphocytes to UV-B and solar irradiation. *Cancer Res* 1993;53:609-14.
32. Alapetite C, Wachter T, Sage E, Moustacchi E. Use of the alkaline comet assay to detect DNA repair deficiencies in human fibroblasts exposed to UVC, UVB, UVA and gamma-rays. *Int J Radiat Biol* 1996;69:359-69.
33. Freeman SE, Hacham H, Gange RW, Maytum DJ, Sutherland JC, Sutherland BM. Wavelength dependence of pyrimidine dimer formation in DNA of human skin irradiated in situ with ultraviolet light. *Proc Natl Acad Sci U S A* 1989;86:5605-9.
34. Ley RD. Ultraviolet radiation A-induced precursors of cutaneous melanoma in *Monodelphis domestica*. *Cancer Res* 1997;57:3682-4.
35. Burren R, Scaletta C, Frenk E, Panizzon RG, Applegate LA. Sunlight and carcinogenesis: expression of p53 and pyrimidine dimers in human skin following UVA I, UVA I + II and solar simulating radiations. *Int J Cancer* 1998;76:201-6.
36. Murphy GM, Norris PG, Young AR, Corbett MF, Hawk JL. Low-dose ultraviolet-B irradiation depletes human epidermal Langerhans cells. *Br J Dermatol* 1993;129:674-7.
37. Hersey P, Bradley M, Hasic E, Haran G, Edwards A, McCarthy WH. Immunological effects of solarium exposure. *Lancet* 1983;1:545-8.
38. Beissert S, Ullrich SE, Hosoi J, Granstein RD. Supernatants from UVB radiation-exposed keratinocytes inhibit Langerhans cell presentation of tumor-associated antigens via IL-10 content. *J Leukoc Biol* 1995;58:234-40.
39. Rivas JM, Ullrich SE. Systemic suppression of delayed-type hypersensitivity by supernatants from UV-irradiated keratinocytes: an essential role for keratinocyte-derived IL-10. *J Immunol* 1992;149:3865-71.
40. Chomarat P, Risoan MC, Banchereau J, Miossec P. Interferon gamma inhibits interleukin 10 production by monocytes. *J Exp Med* 1993;177:523-7.

41. Iwai I, Hatao M, Naganuma M, Kumano Y, Ichihashi M. UVA-induced immune suppression through an oxidative pathway. *J Invest Dermatol* 1999;112:19-24.
42. Bestak R, Halliday GM. Chronic low-dose UVA irradiation induces local suppression of contact hypersensitivity, Langerhans cell depletion and suppressor cell activation in C3H/HeJ mice. *Photochem Photobiol* 1996;64:969-74.
43. Reeve VE, Bosnic M, Boehm-Wilcox C, Nishimura N, Ley RD. Ultraviolet A radiation (320-400 nm) protects hairless mice from immunosuppression induced by ultraviolet B radiation (280-320 nm) or cis-urocanic acid. *Int Arch Allergy Immunol* 1998;115:316-22.
44. Damian DL, Barnetson RS, Halliday GM. Low-dose UVA and UVB have different time courses for suppression of contact hypersensitivity to a recall antigen in humans. *J Invest Dermatol* 1999;112:939-44.
45. LeVee GJ, Oberhelman L, Anderson T, Koren H, Cooper KD. UVA II exposure of human skin results in decreased immunization capacity, increased induction of tolerance and a unique pattern of epidermal antigen-presenting cell alteration. *Photochem Photobiol* 1997;65:622-9.
46. Fourtanier A, Gueniche A, Compan D, Walker SL, Young AR. Improved protection against solar-simulated radiation-induced immunosuppression by a sunscreen with enhanced ultraviolet A protection. *J Invest Dermatol* 2000;114:620-7.
47. Sjoval P, Christensen OB. Local and systemic effect of ultraviolet irradiation (UVB and UVA) on human allergic contact dermatitis. *Acta Derm Venereol* 1986;66:290-4.
48. Mukherji B, Chakraborty NG. Immunobiology and immunotherapy of melanoma [see comments]. *Curr Opin Oncol* 1995;7:175-84.
49. Nestle FO, Aljagic S, Gilliet M, Sun Y, Grabbe S, Dummer R, et al. Vaccination of melanoma patients with peptide- or tumor lysate-pulsed dendritic cells [see comments]. *Nat Med* 1998;4:328-32.
50. Rosenberg SA, Yang JC, Schwartzentruber DJ, Hwu P, Marincola FM, Topalian SL, et al. Immunologic and therapeutic evaluation of a synthetic peptide vaccine for the treatment of patients with metastatic melanoma [see comments]. *Nat Med* 1998;4:321-7.
51. Ceballos PI, Ruiz-Maldonado R, Mihm MC Jr. Melanoma in children [see comments]. *N Engl J Med* 1995;332:656-62.
52. Tucker MA, Misfeldt D, Coleman CN, Clark WH Jr, Rosenberg SA. Cutaneous malignant melanoma after Hodgkin's disease. *Ann Intern Med* 1985;102:37-41.
53. Greene MH, Young TI, Clark WH Jr. Malignant melanoma in renal-transplant recipients. *Lancet* 1981;1:1196-9.
54. Veness MJ, Quinn DJ, Ong CS, Keogh AM, Macdonald PS, Cooper SG, et al. Aggressive cutaneous malignancies following cardiothoracic transplantation: the Australian experience. *Cancer* 1999;85:1758-64.
55. Swerdlow AJ, Weinstock MA. Do tanning lamps cause melanoma? An epidemiologic assessment. *J Am Acad Dermatol* 1998;38:89-98.
56. Spencer JM, Amonette RA. Indoor tanning: risks, benefits, and future trends. *J Am Acad Dermatol* 1995;33:288-98.
57. Gies HP, Roy CR, Elliott G. Artificial suntanning: spectral irradiance and hazard evaluation of ultraviolet sources. *Health Phys* 1986;50:691-703.
58. Larko O, Diffey BL. Natural UV-B radiation received by people with outdoor, indoor, and mixed occupations and UV-B treatment of psoriasis. *Clin Exp Dermatol* 1983;8:279-85.
59. Chen YT, Dubrow R, Zheng T, Barnhill RL, Fine J, Berwick M. Sunlamp use and the risk of cutaneous malignant melanoma: a population-based case-control study in Connecticut, USA. *Int J Epidemiol* 1998;27:758-65.
60. Walter SD, King WD, Marrett LD. Association of cutaneous malignant melanoma with intermittent exposure to ultraviolet radiation: results of a case-control study in Ontario, Canada. *Int J Epidemiol* 1999;28:418-27.
61. Westerdahl J, Ingvar C, Masback A, Jonsson N, Olsson H. Risk of cutaneous malignant melanoma in relation to use of sunbeds: further evidence for UV-A carcinogenicity. *Br J Cancer* 2000;82:1593-9.
62. Diffey BL, Farr PM. Tanning with UVB or UVA: an appraisal of risks. *J Photochem Photobiol B* 1991;8:219-23.
63. Stern RS, Nichols KT, Vakeva LH. Malignant melanoma in patients treated for psoriasis with methoxsalen (psoralen) and ultraviolet A radiation (PUVA): the PUVA Follow-Up Study [see comments]. *N Engl J Med* 1997;336:1041-5.
64. Holman CD, Armstrong BK, Heenan PJ. Relationship of cutaneous malignant melanoma to individual sunlight-exposure habits. *J Natl Cancer Inst* 1986;76:403-14.
65. Evans RD, Kopf AW, Lew RA, Rigel DS, Bart RS, Friedman RJ, et al. Risk factors for the development of malignant melanoma: I: review of case-control studies. *J Dermatol Surg Oncol* 1988;14:393-408.
66. Naylor MF, Boyd A, Smith DW, Cameron GS, Hubbard D, Neldner KH. High sun protection factor sunscreens in the suppression of actinic neoplasia. *Arch Dermatol* 1995;131:170-5.
67. Thompson SC, Jolley D, Marks R. Reduction of solar keratoses by regular sunscreen use [see comments]. *N Engl J Med* 1993;329:1147-51.
68. Green A, Williams G, Neale R, Hart V, Leslie D, Parsons P, et al. Daily sunscreen application and betacarotene supplementation in prevention of basal-cell and squamous-cell carcinomas of the skin: a randomised controlled trial [published erratum appears in *Lancet* 1999 Sep 18;354(9183):1038] [see comments]. *Lancet* 1999;354:723-9.
69. Klepp O, Magnus K. Some environmental and bodily characteristics of melanoma patients: a case-control study. *Int J Cancer* 1979;23:482-6.
70. Graham S, Marshall J, Haughey B, Stoll H, Zielezny M, Brasure J, et al. An inquiry into the epidemiology of melanoma. *Am J Epidemiol* 1985;122:606-19.
71. Beitner H, Norell SE, Ringborg U, Wennersten G, Mattson B. Malignant melanoma: aetiological importance of individual pigmentation and sun exposure. *Br J Dermatol* 1990;122:43-51.
72. Autier P, Dore JF, Schiffiers E, Cesarini JP, Bollaerts A, Koelmel KF, et al. Melanoma and use of sunscreens: an EORTC case-control study in Germany, Belgium and France. The EORTC Melanoma Cooperative Group. *Int J Cancer* 1995;61:749-55.
73. Westerdahl J, Olsson H, Masback A, Ingvar C, Jonsson N. Is the use of sunscreens a risk factor for malignant melanoma? *Melanoma Res* 1995;5:59-65.
74. Wolf P, Quehenberger F, Mullegger R, Stranz B, Kerl H. Phenotypic markers, sunlight-related factors and sunscreen use in patients with cutaneous melanoma: an Austrian case-control study. *Melanoma Res* 1998;8:370-8.
75. Osterlind A, Tucker MA, Stone BJ, Jensen OM. The Danish case-control study of cutaneous malignant melanoma. II. Importance of UV-light exposure. *Int J Cancer* 1988;42:319-24.
76. Elwood JM, Gallagher RP. More about: sunscreen use, wearing clothes, and number of nevi in 6- to 7-year-old European children [letter; comment]. *J Natl Cancer Inst* 1999;91:1164-6.
77. Holly EA, Aston DA, Cress RD, Ahn DK, Kristiansen JJ. Cutaneous melanoma in women. I. Exposure to sunlight, ability to tan, and other risk factors related to ultraviolet light. *Am J Epidemiol* 1995;141:923-33.
78. Rodenas JM, Delgado-Rodriguez M, Herranz MT, Tercedor J, Serrano S. Sun exposure, pigmentary traits, and risk of cutaneous malignant melanoma: a population-based case-control study in Spain. *Int J Epidemiol* 2000;29:103-10.

- neous malignant melanoma: a case-control study in a Mediterranean population [see comments]. *Cancer Causes Control* 1996;7:275-83.
79. Arranz JE, Hernandez JJS, Fernandez PB, Gonzalez-Baron M, Aunon PX, Arranz EE, et al. Cutaneous malignant melanoma and sun exposure in Spain. *Melanoma Res* 1999;9:199-205.
 80. Autier P, Dore JF, Negrier S, Li nard D, Panizzon R, Lejeune FJ, et al. Sunscreen use and duration of sun exposure: a double-blind, randomized trial. *J Natl Cancer Inst* 1999;91:1304-9.
 81. Kaidbey KH, Barnes A. Determination of UVA protection factors by means of immediate pigment darkening in normal skin. *J Am Acad Dermatol* 1991;25:262-6.
 82. Garland CF, Garland FC, Gorham ED. Could sunscreens increase melanoma risk? *Am J Public Health* 1992;82:614-5.
 83. Garland CF, Garland FC, Gorham ED. Re: Effect of sunscreens on UV radiation-induced enhancement of melanoma growth in mice [letter; comment]. *J Natl Cancer Inst* 1994;86:798-800.
 84. Beral V, Evans S, Shaw H, Milton G. Malignant melanoma and exposure to fluorescent lighting at work. *Lancet* 1982;2:290-3.
 85. Rigel DS, Friedman RJ, Levenstein MJ, Greenwald DI. Relationship of fluorescent lights to malignant melanoma: another view. *J Dermatol Surg Oncol* 1983;9:836-8.
 86. Sorahan T, Grimley RP. The aetiological significance of sunlight and fluorescent lighting in malignant melanoma: a case-control study. *Br J Cancer* 1985;52:765-9.
 87. English DR, Rouse IL, Xu Z, Watt JD, Holman CD, Heenan PJ, et al. Cutaneous malignant melanoma and fluorescent lighting. *J Natl Cancer Inst* 1985;74:1191-7.
 88. Swerdlow AJ, English JS, MacKie RM, O'Doherty CJ, Hunter JA, Clark J, et al. Fluorescent lights, ultraviolet lamps, and risk of cutaneous melanoma [published erratum appears in *BMJ* 1988 Nov 5;297(6657):1172]. *BMJ* 1988;297:647-50.
 89. Morison WL. Phototherapy and photochemotherapy: an update. *Semin Cutan Med Surg* 1999;18:297-306.
 90. Rhodes AR, Harfist TJ, Momtaz TK. The PUVA-induced pigmented macule: a lentiginous proliferation of large, sometimes cytologically atypical, melanocytes. *J Am Acad Dermatol* 1983;9:47-58.
 91. Konrad K, Gschnalt F, Wolff K. Ultrastructure of poikiloderma-like pigmentary changes after repeated experimental PUVA-overdosage. *J Cutan Pathol* 1977;4:219-20.
 92. Miller R. Psoralens and UV-A-induced stellate hyperpigmented freckling. *Arch Dermatol* 1982;118:619-20.
 93. Szekeres E, Torok L, Szucs M. Auftreten disseminierter hyperpigmentierter frecke unter PUVA-behandlung. *Hautarzt* 1981;32:33-5.
 94. Zelickson AS, Mottaz JH, Muller SA. Melanocyte changes following PUVA therapy. *J Am Acad Dermatol* 1979;1:422-30.
 95. Hashimoto K, Kohda H, Kumakiri M, Blender SL, Willis I. Psoralen-UVA-treated psoriatic lesions: ultrastructural changes. *Arch Dermatol* 1978;114:711-22.
 96. Abel EA, Reid H, Wood C, Hu CH. PUVA-induced melanocytic atypia: is it confined to PUVA lentiginos? *J Am Acad Dermatol* 1985;13:761-8.
 97. Bleehen SS. Freckles induced by PUVA treatment [proceedings]. *Br J Dermatol* 1978;99:20.
 98. Cox NH, Jones SK, Downey DJ, Tuyp EJ, Jay JL, Moseley H, et al. Cutaneous and ocular side-effects of oral photochemotherapy: results of an 8-year follow-up study. *Br J Dermatol* 1987;116:145-52.
 99. Roth DE, Hodge SJ, Callen JP. Possible ultraviolet A-induced lentiginos: a side effect of chronic tanning salon usage. *J Am Acad Dermatol* 1989;20:950-4.
 100. Jones SK, Moseley H, MacKie RM. UVA-induced melanocytic lesions. *Br J Dermatol* 1987;117:111-5.
 101. Robbins JH, Moshell AN. DNA repair processes protect human beings from premature solar skin damage: evidence from studies on xeroderma pigmentosum. *J Invest Dermatol* 1979;73:102-7.
 102. Hannuksela-Svahn A, Sigurgeirsson B, Pukkala E, Lindelof B, Berne B, Hannuksela M, et al. Trioxsalen bath PUVA did not increase the risk of squamous cell skin carcinoma and cutaneous malignant melanoma in a joint analysis of 944 Swedish and Finnish patients with psoriasis. *Br J Dermatol* 1999;141:497-501.
 103. Lindelof B, Sigurgeirsson B, Tegner E, Larko O, Johannesson A, Berne B, et al. PUVA and cancer risk: the Swedish follow-up study. *Br J Dermatol* 1999;141:108-12.
 104. Naylor MF. Erythema, skin cancer risk, and sunscreens [editorial]. *Arch Dermatol* 1997;133:373-5.
 105. Wulf HC, Poulsen T, Brodthagen H, Hou-Jensen K. Sunscreens for delay of ultraviolet induction of skin tumors. *J Am Acad Dermatol* 1982;7:194-202.
 106. Moan J, Dahlback A, Setlow RB. Epidemiological support for an hypothesis for melanoma induction indicating a role for UVA radiation. *Photochem Photobiol* 1999;70:243-7.

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